

Molecular identification of *Pleurotus ostreatus* strains from Patagonia and their ability to retain laccase activity at low temperature

Identificación molecular de cepas de *Pleurotus ostreatus* de la Patagonia y su capacidad para retener la actividad lacasa a baja temperatura

Maximiliano Rugolo ¹

Francisco Kuhar ¹

Maria Belén Pildain ¹

Bernardo E. Lechner ²

Mario Rajchenberg ¹

¹ National Research Council of Argentina (CONICET), Centro de Investigación y Extensión Forestal Andino Patagónico, C.C. 14, 9200 Esquel, Chubut, Argentina and Universidad Nacional de la Patagonia S.J. Bosco, Ruta 259 km 16.4, 9200 Esquel, Chubut, Argentina

² CONICET / Universidad de Buenos Aires, Instituto de Micología y Botánica (InMiBo), Departamento de Biodiversidad y Biología Experimental, Facultad de Ciencias Exactas y Naturales.

Maximiliano Rugolo, e-mail: maxirugolo@gmail.com

ORCID: 0000-0003-2090-9791

Resumen

Antecedentes: *Pleurotus* spp. son hongos reconocidos por su capacidad de degradar biológicamente materiales lignocelulósicos. En Argentina se conocen 6 especies de *Pleurotus* nativas, siendo *P. pulmonarius* y *P. ostreatus* las de mayor interés biotecnológico.

Objetivo: Determinar las relaciones filogenéticas de cepas de *Pleurotus* de la Patagonia y su capacidad enzimática (lacasa) a bajas temperaturas.

Metodología: Se realizó un análisis filogenético de las regiones ITS de 8 cepas de *Pleurotus* de Patagonia y la región de Paraná en el noreste y centro de la Argentina. Para cuantificar la actividad lacasa se usó dimetoxifenol (DMP) como sustrato, incubando a 30 y 10 °C, con y sin tratamiento previo de calor a 80 °C.

Resultados y conclusiones: El análisis filogenético determinó que las cepas provenientes de la Patagonia (5) correspondían a *P. ostreatus*, y las no patagónicas a *P. pulmonarius* (3). Con respecto a la actividad de lacasa, no se observó una relación entre la capacidad de retención y el origen de las cepas, registrándose más del 50 % de retención de la actividad a 10 °C, por lo que su uso en procesos biotecnológicos que requieren condiciones de bajas temperaturas es potencialmente viable.

Palabras clave: *Araucaria* spp., enzimas adaptadas al frío, hongos ostra

Abstract

Background: *Pleurotus* spp. are fungi characterized by their ability to biologically degrade lignocellulosic materials. In Argentina, 6 wild *Pleurotus* species are known, being *P. pulmonarius* and *P. ostreatus* the ones with the greatest biotechnological interest.

Objective: To determine the phylogenetic relationships of *Pleurotus* strains from Patagonia and their enzymatic capacity (laccase) at low temperatures.

Methods: A phylogenetic analysis of the ITS regions of 8 wild *Pleurotus* strains from Patagonia and Parana region in northeast and central Argentina was carried out. Laccase activity of the strains was quantified using dimethoxyphenol (DMP) as substrate, incubating at 30 and 10 °C, with and without previous heat treatment at 80 °C.

Results and conclusions: Phylogenetic analysis of the ITS region of *Pleurotus* strains showed that Patagonian strains belong to *P. ostreatus* (5) and non-Patagonian strains to *P. pulmonarius* (3). Laccase activity was no relationship between the retention capacity and the origin of the strains, with more than 50% retention of activity at 10 °C, therefore it viable their use in biotechnological processes that require low temperature conditions.

Key words: *Araucaria* spp., cold-adapted enzymes, oyster mushroom

Received: 17/07/2018

Accepted: 03/06/2019

Published on line: 17/07/2019

Introduction

The genus *Pleurotus* in Argentina is known mainly from morphological studies. Lechner *et al.* (2004) concluded that there are, so far, six species: *P. albidus* (Berk.) Pegler, *P. cystidiosus* O. K. Mill., *P. ostreatus* (Jacq.) P. Kumm., *P. pulmonarius* Fr. (Quél.), *P. rickii* Bres. and *P. djamor* (Rumph. ex Fr.) Boedijn (with three varieties: *djamor*, *cyathiformis* and *roseus*). The species have been found in the Pampas (growing on *Populus* and *Salix* trunks) and in tropical and subtropical regions (growing on *Araucaria angustifolia* and other tropical woods). *Pleurotus ostreatus* was only found in Patagonia growing on the native host *A. araucana* (Lechner *et al.*, 2002). In Brazil Menolli *et al.* (2014) confirmed the presence of *P. albidus*, *P. djamor*, *P. fuscusquamulosus* D.A. Reid & Eicker, *P. pulmonarius*, and *P. rickii* with nrITS sequences study.

An accurate assessment of the genetic diversity of Argentinian *Pleurotus* wild strains is needed to establish their identities and allow accurate bases for their efficient use.

The white rot fungi *Pleurotus* spp. are characterized by their ability to degrade the lignin polymer of wood tissues. They produce a several oxidative enzymes including lignin peroxidase (LiP), manganese peroxidase (MnP), laccases and oxidases involved in the process (Mata *et al.*, 2017). The laccases, a multicopper oxidases with low substrate specificity and strong oxidative ability (Wang *et al.*, 2010) can be used for a variety of applications including paper pulp bleaching, decolorization of textile dyes, transformation of lignin into its derivatives, and detoxification of harmful substances (Saparrat *et al.*, 2008). The most efficient laccase-producers are white-rot fungi, such as *Trametes versicolor* (L.) Lloyd., *Trametes hirsuta* (Wulfen) Pilát and *Pleurotus ostreatus* (Šnajdr and Baldrian, 2007). For commercial applications, they usually require unique properties such as thermal stability and resistance to acid or alkaline conditions.

Enzyme activities show, in general, a progressive increase with temperatures till they reach the denaturation area, with the consequent (and sometimes irreversible) loss of activity. Coordination environment changes of the metal sites are also responsible for the loss of activity at high temperatures (Bonomo *et al.*, 2001).

An important application of laccases in food industry is must beer and wine stabilization as an alternative to physical-chemical adsorbents (Osma *et al.*, 2010). These complex mixtures of different chemical compounds have to be stabilized at low temperatures, increasing their shelf life, avoiding haze formation and preserving organoleptic madeirization-resistant features. These enzymes could increase productivity, efficiency and quality of food products without a costly investment and has the advantage of being a mild technology (Minussi *et al.*, 2002).

Cold-adapted and thermostable enzymes are needed in industrial processes. Although low temperature conditions do not irreversible inactivate enzymes, their activities decrease to ranges in which they are not adequate for industrial processes (Cavicchioli *et al.*, 2002). Patagonian microorganisms have adaptations to the stringent environmental conditions where they grow (Torres *et al.*, 2016). These adaptations include the production of steroids, ergosterols (Dufourc, 2008), antioxidant compounds, carbohydrates, proteins and enzymes adapted to extreme cold conditions. Laccases of the edible mushrooms of the genus *Pleurotus*, growing naturally in Patagonia, could be used in different biotechnological applications such as food and beverage industries if they prove to provide cold-adapted enzymes.

The objectives of this work were (a) to elucidate the phylogenetic relationships of *Pleurotus* strains from Patagonia as a preliminary step in order to select those that are feasible to use in industrial processes that require cold temperatures and (b) to evaluate the laccase activity at low temperatures. We hypothesize that Patagonian strains retain a higher laccase activity at low temperatures than those from lower latitudes.

Materials and Methods

Strains and culture media

Eight strains of *Pleurotus* spp. were tested; they are kept at CIEFAPcc (Colección de Cultivos, Centro de Investigación y Extensión Forestal Andino Patagónico at the first author address) and at BAFC (Buenos Aires Facultad de Ciencias, Universidad de Buenos Aires Culture Collection). Locations of these strains are shown in Figure 1.

Pleurotus ostreatus, ARGENTINA, Neuquén, Villa Pehuenia, Paso del Arco a trail from Villa Pehuenia City to the international limit Paso del Arco with more than 50 kilometers

from 1100 to 1400 msnm, leg. *M. Rugolo & M. Rajchenberg*, May 3, 2013, CIEFAPcc 617, 621, 622, y 623; Circuito Pehuenia, Moquehue, a trail from Alumine City to Villa Pehuenia City with more than 100 kilometers (from 900 to 1300 msnm) leg. *J. del Vas*, March 3, 1994, CIEFAPcc 104. Growing on *Araucaria araucana*.

Pleurotus ostreatus, ARGENTINA, Chubut, Aldea Escolar, next to Arroyo Blanco, leg. *M. Rugolo*, June 26, 2013, CIEFAPcc 616. Growing on *Populus* sp.

Pleurotus pulmonarius, ARGENTINA, Buenos Aires, Zárate, leg. *B. Lechner*, May 10, 2011, BAFC 4281. Growing on *Populus* sp.

Pleurotus pulmonarius, ARGENTINA, Misiones, San Pedro, leg. *E. Albertó*, May 27, 2001, BAFC 263. Growing on *Araucaria angustifolia*.

The culture media used for micelial growth was agar malt extract (Difco). Liquid cultures were prepared with 12.5 g of malt extract in 1 l distilled water in 250 mL flasks. The mycelium was incubated in static conditions at 23 °C in darkness (Rugolo, 2018). Supernatant was harvested after 30 days, when the colonization was completed.

PRE PRINT VERSION

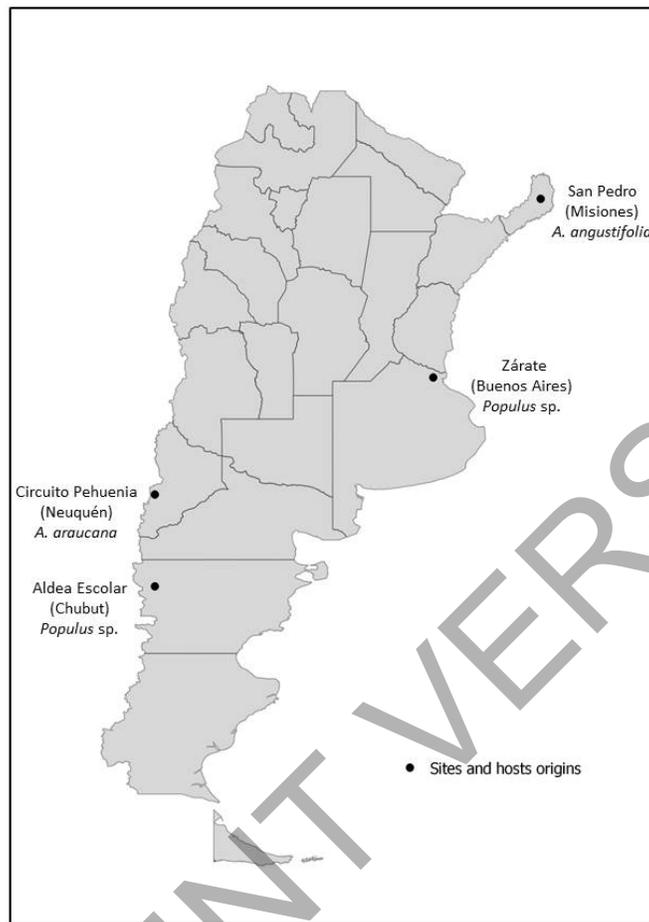


Figure 1. Map of Argentina. Locations and hosts of different strains tested.

Molecular determination

DNA extraction and PCR conditions. DNA was extracted from freshly collected mycelium from pure culture grown in liquid malt peptone broth with 10% (v/v) of malt extract (Merck) and 0.1% (w/v) Bacto peptone (Difco), in 15 mL tubes at 24°C in the dark. DNA extractions were carried out with the UltraClean™ Microbial DNA Isolation Kit (MO BIO Laboratories Inc., Solana Beach, California), following the manufacturers protocols. PCR for the full Internal Transcribed Spacer region (i.e., ITS1, ITS2 and the intervening 5.8S RNA gene; further referred as ITS) amplified with the primers ITS1

(TCCGTA GGTGAACCT) -ITS4 (TCCTCCGCTTATTGATATGC) (White et al 1990). The PCR conditions were described by Imtiaz *et al.* (2011): 95 °C for 2 min, 30 cycles of 94 °C for 30 s, 51 °C for 30 s, 72 °C for 1 min, followed by 72 °C for 10 min. The amplified fragments were purified and sequenced at the DNA Synthesis and Sequencing Facility, Macrogen (Seoul, South Korea). Sequences generated in this study were submitted to GenBank (MK421536 to MK421541 and MK530228-MK530229).

Sequence and phylogenetic analyses.“Blastn” of the obtained sequences were performed against Genbank database (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>). Available ITS sequences of the *P. ostreatus* group obtained from previous studies (Vilgalys *et al.*, 1996; Huerta *et al.*, 2010; Menolli *et al.*, 2014) were included. No outgroup was chosen, the tree being rooted to midpoint.

Nucleotide sequences for the ITS region were initially edited with BioEdit 7.0.9.0 (Hall, 1999), then aligned using L-INS-i strategy as implemented in MAFFT v 7.0 (Katoh and Standley, 2013) and manually adjusted using MEGA version 6 (Tamura *et al.*, 2013). The final ITS dataset comprised 40 sequences and 569 characters including gaps. The best-fit models of evolution were determined using the AIC criterion (Akaike, 1974) implemented in jModelTest (Posada, 2008) and was GTR+G. Phylogenetic analysis of the data set was performed using Maximum likelihood (ML) analysis was performed in PHYML executed on the south of France bioinformatics platform (<http://www.atgc-montpellier.fr/phyml/>) following Guindon *et al.*, (2010) under the GTR nucleotide substitution model. Bootstrap values of the most likely tree were calculated with 1000 repetitions.

Laccase activity assays with *Pleurotus* strains

Laccase activity (E.C.: 1.10.3.2) was measured in supernatant using 2,6-dimethoxyphenol (DMP) 5 mM in 0.1 M sodium acetate buffer (pH 3.6) at 30 °C, 10 °C, and after heat treatment (the supernatant was exposed to 80 °C, in thermal bath, for 20 min). Oxidation of DMP was determined by the increase in A469 ($\epsilon_{469}=27 \text{ mM}\cdot\text{cm}^{-1}$) according Paszczynski and Crawford (1991). Retention of laccase activity at low temperatures was calculated as:

$$\left[\frac{\text{activity measured at } 10 \text{ }^\circ\text{C}}{\text{activity measured at } 30 \text{ }^\circ\text{C}} \right] * 100$$
 (Yuan *et al.*, 2016). Thermal stability was calculated as the percentage of remaining activity after thermal treatment measured at 30 °C. The measurements consisted in three replicates. Analysis of variance (ANOVA) were used with the measures of EU and the significant differences between treatments were compared by Tukey’s test at 5% level of probability. The statistical treatments were tested by the software InfoStat software (Di Rienzo *et al.*, 2011). Enzyme activity was expressed in International Enzymatic Units (EU) as the amount of enzyme required to release 1 μmol of product in 1 min.

Results and Discussion

Molecular identification

ITS analyses including 40 taxa inferred from Maximum Likelihood (ML) confirmed that all strains from Patagonia clustered within *P. ostreatus* species (94% Bootstrap value), while the strains isolated from center and NE Argentina belong to *P. pulmonarius* (89%) (Figure 2). The latter has concordance with the tropical and subtropical distribution of that taxon as reported by Lechner *et al.*, (2004) and Menolli *et al.* (2014).

The description of Patagonian species based on macro- and microscopical features match with *P. pulmonarius*, but interespecific mating tests showed full compatibility with *P. ostreatus* (Lechner *et al.* 2002). Here we confirm the identity of the Patagonian strains as *Pleurotus ostreatus*.

According to Vilgalys (1997) the complex “*Pleurotus ostreatus*” groups *P. ostreatus*, *P. pulmonarius*, *P. eryngii*, *P. abieticola*, and *P. populinus*, which share a “pleurotoid” stature: more or less shelf-like and radically eccentrically stipitate (Albertó *et al.*, 2002). In our phylogenetic reconstruction (Figure 2) we agree with Albertó *et al.* (2002). *Pleurotus* phylogeny is currently based mainly upon ITS; this rDNA region solves the species relationships within the group but the branching pattern within *P. ostreatus* remains poorly determined. Liu *et al.* (2012) demonstrated that RPB2 gene to be superior to ITS and EF1 α for distinguishing Chinese *P. ostreatus*. For a complete understanding of the genetic diversity of this species, including more Patagonian strains, future studies must include a multilocus approach using more variable gene regions and/or genotyping in order to assess population diversity.

It is a rarity that both species, *P. ostreatus* and *P. pulmonarius*, are found associated with conifers like *Araucaria araucana* and *Araucaria angustifolia*, respectively. The nature of the substrate points out towards the use of these strains for futures assays to evaluate growth and production of *Pleurotus* in resinous forest residues.

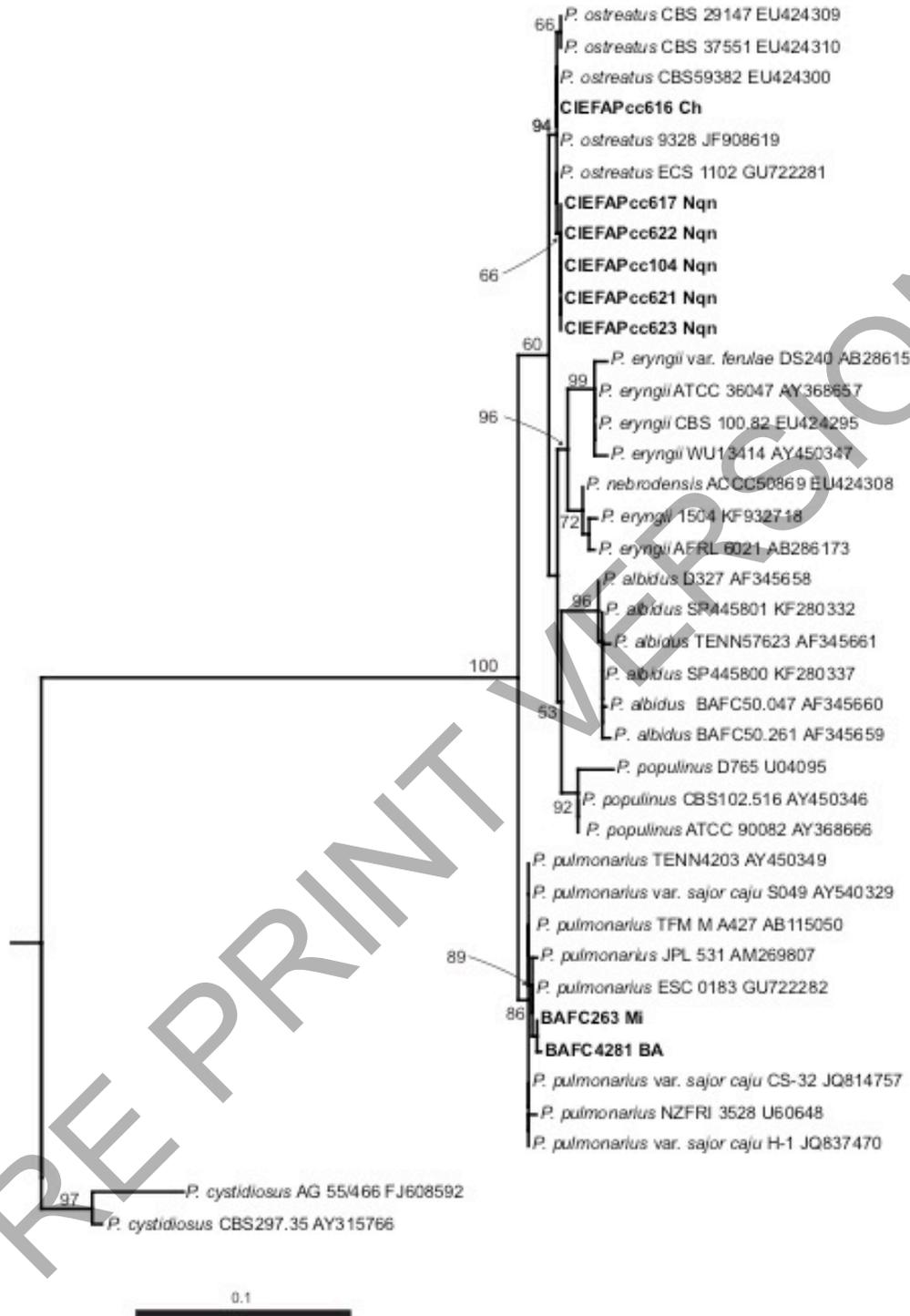


Figure 2. Phylogenetic tree of *Pleurotus* species based on rDNA-ITS sequences using Maximum Likelihood analysis. Numbers on branches indicate the bootstrap values obtained from 1000 replications. Branches supported by bootstrap value greater than 50% are shown. CH, Chubut Province; Nqn, Neuquen Province; MI, Misiones Province; BA, Buenos Aires Province. Sequences in bold were generated in this study.

Laccase activity assays with *Pleurotus* strains

Strains from Neuquén presented the highest laccase activities with values close to 60 EU.L⁻¹ for CIEFAPcc 104, 617, 621 and 622 (Figure 3). Strains of *P. ostreatus* 622 and 616, and strain of *P. pulmonarius* 263 and 4281 showed the highest retention of laccase activity at 10 °C, with 70 %, 69 %, 58 % and 60 %, respectively (Figure 4), in comparison with their activity at 30 °C. Strain 623 presented the highest thermal stability after treatment at 80 °C (Figure 5).

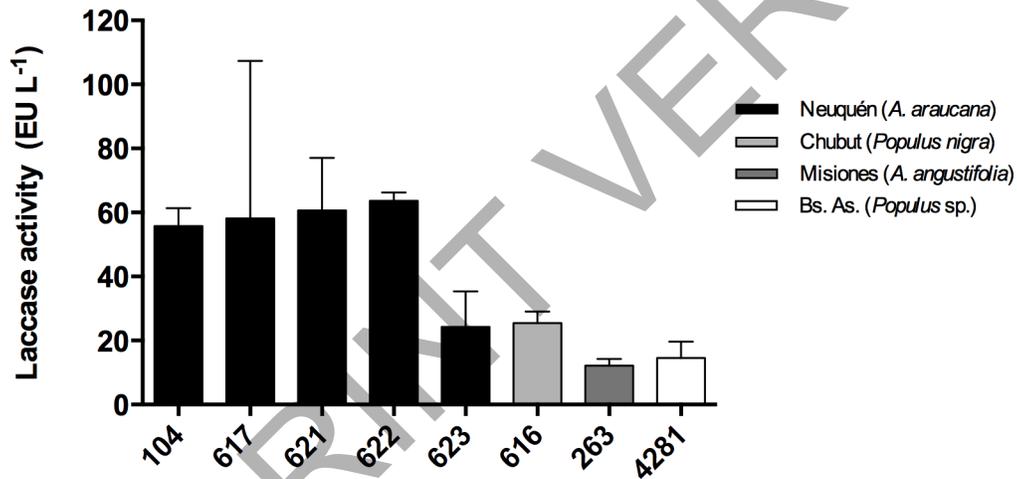


Figure 3. Laccase activity of the strains at 30 °C. Different numbers correspond to different *Pleurotus ostreatus* strains. Colors indicate the geographical origin of each strain. Means with the same letter are not significantly different ($p > 0.05$).

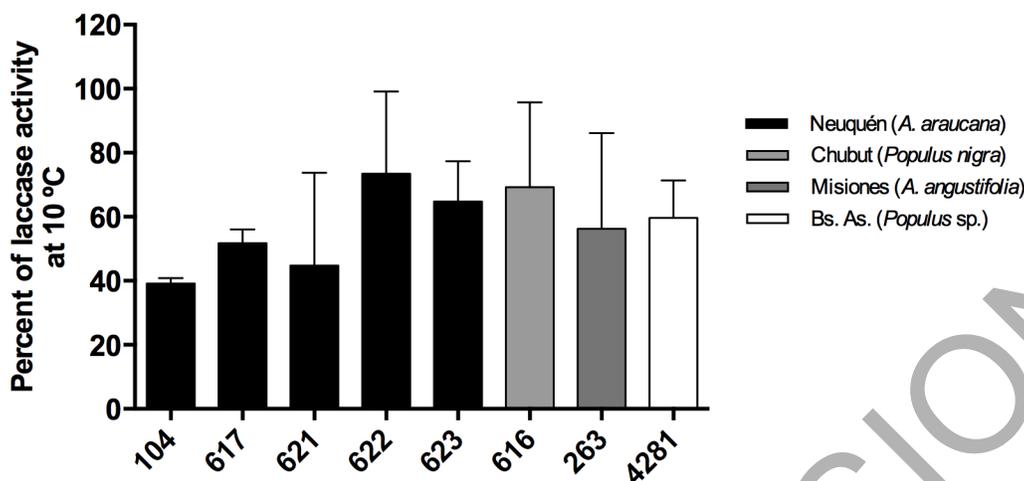


Figure 4. Percent retention of laccase activity at 10 °C. Different numbers correspond to different *Pleurotus ostreatus* strains. Colors indicate the geographical origin of each strain. Means with the same letter are not significantly different ($p > 0.05$).

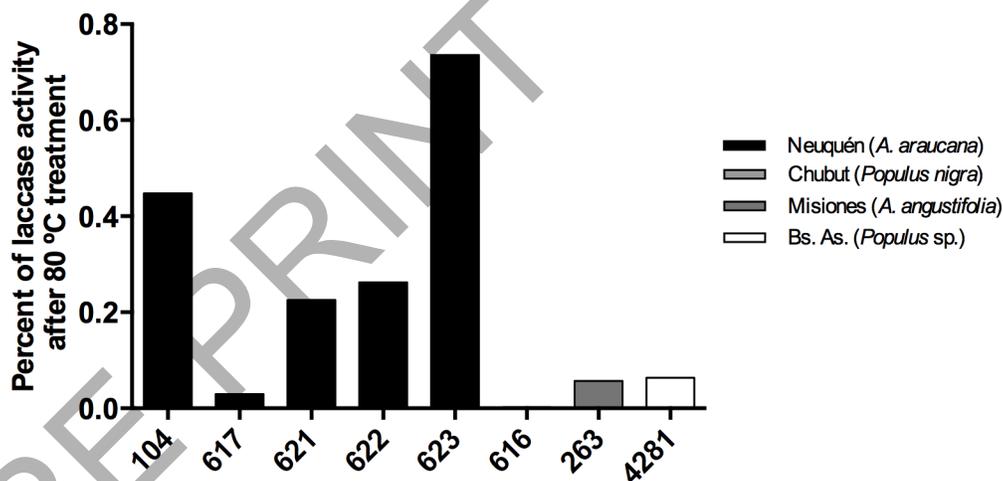


Figure 5. Thermostability of laccase activity at 30 °C, after heat treatment. Different numbers correspond to different *Pleurotus ostreatus* strains. Colors indicate the geographical origin of each strain.

Wang *et al.* (2010) showed better results with a *Pycnoporus* strain (from Jiangsu, China), with two purified stable laccases in a range of 0-50 °C, and a remaining half-activity for 40 days at 30 °C. When temperature exceeded 60 °C *Pycnoporus* enzymes activities decreased

rapidly, and they were inactivated after 1 incubation h at 80 °C. Thermostability of the three laccase isoforms from *Pleurotus nebrodensis* was assayed by Yuan *et al.* (2016). They showed that Lac2 was the most thermostable and Lac1 demonstrated better thermostability compared with Lac3 in the temperature range from 30 °C to 70 °C. After exposure Lac2 to 50 °C for 120 min, 70% of its original activity remained. A higher temperature of 70 °C, however, caused rapid inactivation (60% and 80% activity loss within 30 min and 60 min, respectively). Lac3 was completely inactivated at 70 °C after 40 min.

In comparison with our results, those purified laccases were more stable than those from *P. ostreatus* and *P. pulmonarius* strains from Argentina tested in this work. It should be noted that we worked with a total supernatant, in contrast with the purified laccases of *Pycnoporus* SYBC-L1 strain by Wang *et al.* (2010) and *Pleurotus nebrodensis* by Yuan *et al.* (2016). The total supernatant has a commercial benefit for industrial application, because it requires an enzymatic crude supernatant, for example, to precipitate phenolic compounds creating a completely clear juice, or without compromising the quality of beer or wine (Novozymes®).

Šnajdr and Baldrian (2007) showed the optimal laccase activity for a *P. ostreatus* strain from Czech Republic to be at 30 °C with 24 EU.L⁻¹, undergoing a great decline at low temperatures (0.02 EU.L⁻¹ at 10 °C), which also exhibited only 5-18 % of the activity at the optimum temperature at 10 °C. We have a notable difference with Patagonian *P. ostreatus* strains with more than 15 EU.L⁻¹ at 10 °C (more than 40 % of retention activity in all strains). Hua *et al.* (2018) showed that *Pleurotus tuoliensis* enzyme activity changed in response to cold stress. The transcriptional regulation of laccase and ligninolytic peroxidase genes plays an important role in the fruiting bodies of *Pleurotus tuoliensis* under low temperature induction (4 °C), in which the enzymatic activities and transcription levels decrease significantly. At 13 °C, the expression of laccase and peroxidase genes increased, and seems to play a dominant role during nutrition growth.

Smalas *et al.* (2000) proposed that high enzymatic efficiency at low temperatures is generally accompanied by reduced thermal stability because there is an increase in the molecular flexibility of the enzyme active site. However, in the system studied we observed that thermal stability did not decrease when laccase enzymatic activity at low temperature increased. Our results suggest that the supernatant produced by the studied strains can be used in processes at low temperatures but they also show a high thermal stability.

However, we rejected our hypothesis because there was no significant difference of activities at 10°C between *P. ostreatus* strains from Patagonia and *P. pulmonarius* strains from north Argentina. We expected an association between strains and the environmental conditions. Nevertheless, laccase activity at 30 °C and after thermal treatment was lower in *Pleurotus pulmonarius* (which grows in spring and summer of South Hemisphere) than in *Pleurotus ostreatus* (which fruits in autumn and winter season of South Hemisphere), that showed a similar percentage of activity retention at 10 °C.

However, due to the low size number of strains, we cannot conclude easily on the effects of origin or species and the conclusion are bounded for the studied strains, but are not representative of the biodiversity of Patagonia.

References

- Akaike, H., 1974. A new look at the statistical model identification. *IEEE Transactions on Automatic Control* 19: 716-723.
- Albertó, E., R.H. Petersen, K.W. Hughes, B. Lechner, 2002. Miscellaneous notes on *Pleurotus*. *Persoonia* 18: 55-69.
- Bonomo, R.P., G. Cennamo, R. Purrello, A.M. Santoro, R. Zappala, 2001. Comparison of three fungal laccases from *Rigidoporus lignosus* and *Pleurotus ostreatus*: correlation between conformation changes and catalytic activity. *Journal of Inorganic Biochemistry* 83: 67-75.
- Cavicchioli, R., K.S. Siddiqui, D. Andrews, K.R. Sowers, 2002. Low-temperature extremophiles and their applications. *Current Opinion in Biotechnology* 13: 253-261.
- Di Rienzo, J.A., F. Casanoves, M.G. Balzarini, L. Gonzalez, M. Tablada, C.W. Robledo, 2011. InfoStat Versión 2011. Grupo InfoStat, Universidad Nacional de Córdoba, Córdoba, Argentina.
- Dufourc, E., 2008. Sterols and membrane dynamics. *Journal of Chemical Biology* 1: 63-77.
- Guindon, S., J.F. Dufayard, V. Lefort, M. Anisimova, W. Hordijk, O. Gascuel, 2010. New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. *Systematic Biology* 59: 307-321.
- Hall, T.A., 1999. Bio-Edit: an user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series* 41: 95-98.
- Hua, S., B. Zhang, Y. Fu, B. Qi, Y. Li, F. Tian, Y. Li, 2018. Enzymatic gene expression by *Pleurotus tuoliensis* (Bailinggu): differential regulation under low temperature induction conditions. *World Journal of Microbiology and Biotechnology* 34: 160. DOI: 10.1007/s11274-018-2487-7
- Huerta, G., D. Martinez-Carrera, J.E. Sanchez, H. Leal-Lara, R. Vilgalys, 2010. Genetic relationships between Mexican species of *Pleurotus* analyzing the ITS-region from rDNA. *Micologia Aplicada International* 22: 15-25.
- Imtiaj, A., T.S Lee, S. Ohga, 2011. Sequence variation of *Pleurotus* species collected from Eastern Asia. *Micologia Aplicada International* 23: 1-10.
- Katoh, K., D.M. Standley, 2013. MAFFT Multiple Sequence Alignment Software Version 7: Improvements in Performance and Usability. *Molecular Biology and Evolution* 30: 772-780.
- Lechner, B.E., R. Petersen, M. Rajchenberg, E. Albertó, 2002. Presence of *Pleurotus ostreatus* in Patagonia, Argentina. *Revista Iberoamericana de Micología* 19: 111-114.

Lechner, B.E., J. Wright, E. Albertó, 2004. The genus *Pleurotus* in Argentina. *Mycologia* 96: 845-858.

Liu, Y., S. Wang, Y. Yin, F. Xu, 2012. Evaluation of genetic diversity of Chinese *Pleurotus ostreatus* cultivars using DNA sequencing technology. *Annals of Microbiology* 63: 571-576.

Mata, G., D. Salmones, J.M. Savoie, 2017. Las enzimas lignocelulolíticas de *Pleurotus* spp. In: Sánchez J.E., D.J. Royse (eds.), *La biología, el cultivo y las propiedades nutricionales y medicinales de las setas Pleurotus spp.* El Colegio de la Frontera Sur, Tapachula. Pp. 63-82.

Menolli Jr, N., B.S. Breternitz, M. Capelari, 2014. The genus *Pleurotus* in Brazil: a molecular and taxonomic overview. *Mycoscience* 55: 378-389.

Minussi, R.C., G.M. Pastore, N. Durán, 2002. Potential applications of laccase in the food industry. *Trends in Food Science and Technology* 13: 205-216. DOI 10.1016/S0924-2244(02)00155-3

Osma, J.F., J.L. Toca-Herrera, S. Rodriguez-Couto, 2010. Uses of laccases in the food industry. *Enzyme Research* DOI 10.4061/2010/918761

Paszczynski, A., R.L. Crawford, 1991. Degradation of azo compounds by ligninases from *Phanerochaete chrysosporium* involvement of veratryl alcohol. *Biochemical and Biophysical Research Communication* 178: 1056-63.

Posada, D., 2008 jModelTest: Phylogenetic Model Averaging. *Molecular Biology and Evolution* 25: 1253-1256.

Rugolo, M, 2018. Ensayos de producción, caracterización enzimática y obtención de expolisacáridos de hongos comestibles lignocelulolíticos patagónicos en sustratos provenientes de desechos herbáceos, agrícolas y forestales. Tesis Doctoral, Universidad de Buenos Aires, Facultad de Ciencias Exactas y Naturales. http://hdl.handle.net/20.500.12110/tesis_n6382_Rugolo

Saparrat, M.C.N., P. Mocchiutti, C.S. Liggieri, M.B. Aulicino, N.O. Caffini, P.A. Balatti, 2008. Ligninolytic enzyme ability and potential biotechnology applications of the white-rot fungus *Grammothele subargentea* LPSC n°. 436 strain. *Process Biochemistry* 43: 368-375.

Smalas, A.O., H.K. Leiros, V. Os, N.P. Willassen, 2000. Cold adapted enzymes. *Biotechnology Annual Review* 6: 1-57.

Šnajdr, J., P. Baldrian, 2007. Temperature affects the production, activity and stability of ligninolytic enzymes in *Pleurotus ostreatus* and *Trametes versicolor*. *Folia Microbiologica* 52: 498-502.

Tamura, K., D. Peterson, G. Stecher, M. Nei, S. Kumar, 2011. Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum

parsimony methods. *Molecular Biology and Evolution* 28: 2731-2739.

Torres, S., D. Cajas, G. Palfner, A. Astuya, A. Aballay, C. Pérez, V. Hernández, J. Becerra, 2016. Steroidal composition and cytotoxic activity from fruiting body of *Cortinarius xiphidipus*. *Natural Product Research* DOI: 10.1080/14786419.2016.1185717

Vilgalys, R., 1997. Biodiversity of the oyster mushroom *Pleurotus*. *Mushroom News* 32-35.

Vilgalys, R., J.M. Moncalvo, S.R. Liou, M. Volovsek, 1996. Recent advances in molecular systematics of the genus *Pleurotus*. In: Royse, D.J. (ed), *Mushroom biology and mushroom products*. Pennsylvania State University, University Park. Pp. 91-101.

Wang, Z.X., Y.J. Cai, X.R. Liao, G.J. Tao, Y.Y. Li, F. Zhang, D.B. Zhang, 2010. Purification and characterization of two thermostable laccases with high cold adapted characteristics from *Pycnoporus* sp. SYBC-L1. *Process Biochemistry* 45: 1720-1729.

Yuan, X., G. Tian, Y. Zhao, L. Zhao, H. Wang, T.B. Ng. 2016. Biochemical characteristics of three laccase isoforms from the basidiomycete *Pleurotus nebrodensis*. *Molecules* 21(2), 203. DOI: 10.3390/molecules21020203

PRE PRINT VERSION